

11-06-00

Patent Docket 17341DIV5



IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant: DONOVAN, Stephen) Examiner: Devish Srivastava
) (parent application)
)
Serial Number: pending) Art Unit: 1653 (parent application)
)
Filed: November 2, 2000)
)
For: METHOD FOR TREATING)
THYROID DISORDERS) Irvine, California

NONPROVISIONAL PATENT APPLICATION TRANSMITTAL LETTER

BOX PATENT APPLICATION
Commissioner for Patents
Washington, D.C. 20231

Dear Sir:

Transmitted herein for filing under 35 U.S.C. §121 and 37 C.F.R. 1.53(b) is a new nonprovisional (divisional) patent application for the invention entitled METHOD FOR TREATING THYROID DISORDERS invented by Stephen Donovan. This application is a divisional of prior pending application serial number 09/504,538, filed February 15, 2000.

With regard to the above-identified pending patent application, please find enclosed: (1) this two-page Transmittal Letter; (2) a copy of the 47-page parent patent application, serial number 09/504,538; (3) a copy of the original, executed declaration of the inventor in parent application serial number 09/504,538; (4) a copy of the assignment of the parent patent application serial number 09/504,538 to Allergan Sales, Inc.; (5) a copy of the Information Disclosure Statement and Form PTO 1449 (3 pages) filed in the parent patent application, serial number 09/504,538 (art items not enclosed); (6) a 5 page Preliminary Amendment; and (7) a return postcard.

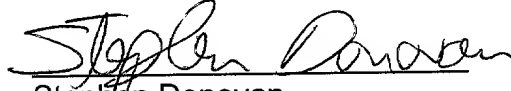
The filing fee is calculated as \$690.00 and should be charged to our deposit account number 01-0885.

Applicant hereby claims the benefit under 35 U.S.C. section 120 from pending application serial number 09/504,538, filed February 15, 2000, the contents of which prior filed application is hereby incorporated by reference in its entirety into the application filed herein.

The pending parent application serial number 09/504,538 is not abandoned by this filing.

The Commissioner is hereby authorized to charge required or necessary fees associated with the filing of this paper and any of the attached documents, and to credit any overpayment to, deposit account number 01-0885.

Date: November 2, 2000


Stephen Donovan
Registration Number 33,433

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CERTIFICATE OF EXPRESS MAILING UNDER 37 C.F.R. § 1.10

I hereby certify that this Nonprovisional Patent Application Transmittal Letter, the specified patent application and the documents referred to as enclosed therein are being deposited with the United States Postal Service on this date November 2, 2000 in an envelope as "Express Mail Post Office to Addressee" Mailing Label number EL385494695US addressed to Box Patent Application, Commissioner for Patents, Washington, D.C. 20231

Stephen Donovan
Name of person mailing paper

Date: November 2, 2000


Signature of person mailing paper

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PRELIMINARY AMENDMENT

Commissioner for Patents
Washington, D.C. 20231

Dear Sir:

Please amend the above-identified patent application as follows:

IN THE SPECIFICATION

1. Please amend the title at page 1, line 1 of the specification from;

"METHOD FOR TREATING THYROID DISORDERS"

to:

--METHOD FOR TREATING HYPOCALCEMIA--.

2. Please add the following at page 1, line 6 (i.e. after the title):

DOCKET # 17341DIV5

--CROSS REFERENCE

This application is a divisional of serial number 09/504,538, filed February 15, 2000--

3. At page 2, line 11 of the specification, please add a comma after the word "States", so that page 2, line 11 reads: "United States, with a lifetime risk of 1%. It is most commonly manifest in the third".

4. At page 7, line 1, please replace the comma after the word "palpation" by the word "and", so that page 7, line 1 reads: " $(\mu\text{Ci/gm})$ /uptake (%). The weight of the thyroid gland is estimated by palpation and".

5. At page 7, line 9 replace the word "potential" by the word --potentially--.

6. Please amend page 7, line 13 from: "thyroid cells by result in later hyperthyroidism" to: --thyroid cells by radiation therapy can result in hypothyroidism--.

7. At page 10, line 27 of the specification, replace "(1976)" with --(1996)--.

8. At page 14, lines 8 and 9, replace both occurrences of the word "Shantz" by --Schantz--.

9. At page 6, line 5 of the specification amend "becoming" to --become--.

10. At page 27, line 8 of the specification before the word "locally" add the words --can be--.

11. At page 29, line 1, delete the second occurrence of the word "known".

12. At page 30, line 27 replace the word "inhibit" by the word –stimulate--.

13. At page 30, line 28 replace the word "follicle" by the word –follicles--.

IN THE CLAIMS

Please amend the claims as follows:

1. (Once amended) A method for treating [a thyroid disorder] hypocalcemia, the method comprising the step of administration of a neurotoxin to a patient, thereby treating [a thyroid disorder] hypocalcemia.

2. Cancel claim 2.

3. Cancel claim 3.

5. Cancel claim 5.

11. (Once amended) A method for treating a [thyroid disorder] hypocalcemia, the method comprising the step of administration of a therapeutically effective amount of a botulinum toxin to a patient, thereby treating [a thyroid disorder] hypocalcemia.

Cancel claims 13-18.

REMARKS

I. Introduction

This application is a divisional application of pending parent application serial number 09/504,538, filed February 15, 2000.

In the parent application serial number 09/504,538, examiner Srivastava set forth during a telephone conversation with the undersigned on August 7, 2000 a six way restriction requirement as follows:

Group I, claims 1-7;

Group II, claims 8-12;

Group III, claims 13-14;

Group IV, claims 15-16.

Group V, claims 17-18, and;

Group VI, claims 19-20.

In response to the restriction requirement in the parent patent application serial number 09/504,538, applicant orally elected with traverse to prosecute Group I, claim 1-7.

The purpose of the present divisional patent application is to now proceed with prosecution of the non-elected and previously cancelled parent application Group VI, claims 19-20.

By this preliminary amendment claims 2-3, 5, and 13-18 have been cancelled, and claims 1 and 11 have been amended. Thus, after entry of this preliminary amendment there are 11 pending claims, these being claims 1, 4, 6-12 and 19-20.

II. Amendments to the Specification

The title of the application has been replaced by a title more descriptive of the claimed subject matter and a cross reference to the parent application has been added after the title.

All of the other amendments made to the specification are made to correct grammatical or typographical errors. The amendment to page 30, line 27 of the specification is supported by at least page 4, lines 24-26 of the specification.

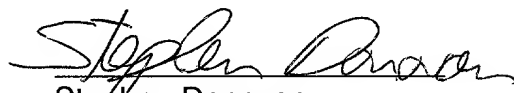
III. Amendments to the Claims

The amendment to claims 1 and 11 are supported by at least the second half of Example 12 on page 41 of the specification and by original claim 19.

IV. Conclusion

Examination and allowance of all claims is requested.

Respectfully Submitted,


Stephen Donovan
Registration Number 33,433

Date: November 2, 2000

Please direct all correspondence and inquiries to:

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METHOD FOR TREATING THYROID DISORDERS

by

Stephen Donovan

BACKGROUND

The present invention relates to methods for treating thyroid disorders. In particular the present invention relates to methods for treating thyroid disorders by administration of a neurotoxin to a patient.

It has been estimated that at least about two hundred million people worldwide are afflicted with a thyroid disorder and women are affected disproportionately, as compared to men, by a ratio of about ten to one. In the United States, about ten million persons, including ten percent of all women over age 45, have either overactive or underactive thyroid glands. Bayliss et al., *Thyroid Disease The Facts*, preface, Oxford University Press (1998).

Thyroid Function

The thyroid is an endocrine gland comprised of follicle cells and non-follicular or C cells. The follicle cells are capable of making two hormones, triiodothyronine (T_3), which contains three iodine atoms and thyroxine (T_4) which contains four. The action of thyroid hormone is concerned principally with the regulation of metabolic rate by, for example, increasing energy production and oxygen consumption by most normal tissues. Synthesis and release of T_3 and T_4 by thyroid cells is influenced by thyroid stimulating hormone (TSH, also called thyrotrophin) made by the pituitary. The C cells can make calcitonin which appears to influence calcium metabolism. Significantly, calcitonin is a potent hypocalcemic agent. Disorders of the thyroid include autoimmune disorders (such as Graves' disease), thyroiditis (inflammation or infection of the thyroid),

and cancer, all of which conditions can result in hypothyroidism (as can occur in Hashimoto's thyroiditis) or hyperthyroidism (thyroidtoxicosis, as can occur in Graves' disease). An enlarged thyroid (goiter) can be euthyroid, or a symptom of either hyperthyroidism (thyroidtoxicosis) or hypothyroidism.

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Most cases of hyperthyroidism are believed to be due to the action of thyroid stimulating antibodies upon the thyroid as a whole (Graves' disease, diffuse toxic goiter).

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Graves' disease has been estimated to occur in 0.4% of the population of the United States with a lifetime risk of 1%. It is most commonly manifest in the third or fourth decade of life and the female to male ratio is about 7:1 to about 10:1.

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The thyroid abnormalities characteristic of Graves' disease apparently result from the action of immunoglobulin of the IgG class on the thyroid. These antibodies may be directed against components or regions of the plasma membrane that include the receptor for thyroid stimulating hormone (TSH) itself. The principal destabilizing factor resulting in autoimmune thyroid disease appears to be an organ specific defect in suppressor T-lymphocytes. Hyperthyroidism itself appears to have an adverse effect on generalized suppressor T-cell function, and this may be a self-perpetuated or potentiating factor in Graves' disease. Significantly, there is no known cure for Grave's disease, treatment being designed merely to reduce the thyroid's ability to produce thyroid hormones.

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Causes of hyperthyroidism besides Graves' disease, include toxic multinodular goiter, toxic adenoma, subacute viral thyroiditis, postpartum thyroiditis, thyroid, gonadal and pituitary tumors and excess pituitary TSH.

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The normal thyroid gland weighs about fifteen grams. It is convex anteriorly and concave posteriorly as a result of its relation to the anterolateral portions of the trachea and larynx, to which it is firmly fixed by fibrous tissue. The two lateral

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lobes of the thyroid extend along the sides of the larynx, reaching the level of the middle of the thyroid cartilage. Each thyroid lobe resides in a bed between the trachea and larynx medially and the carotid sheath and sternocleidomastoid muscles laterally.

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The thyroid is composed of an aggregation of spherical or ovate cystlike follicles of variable size. The interfollicular areas are occupied by a highly vascularized network which includes parafollicular cells (C cells) which are responsible for the secretion of calcitonin. Parathyroid hormone (PTH, made by the parathyroid glands), calcitonin (made by the C cells of the thyroid) and dihydroxycholecalciferol (metabolized from vitamin D in the kidney) are the principal hormones concerned with the metabolism of ions such as calcium, phosphate, pyrophosphate, citrate and magnesium, and with the regulation of the metabolism of bone and its organic constituents. In humans, it is believed that calcitonin acts, in a manner antagonistic to PTH, to lower plasma calcium.

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The thyroid gland is enveloped by a thickened fibrous capsule. The deep cervical fascia divides into an anterior and a posterior sheath, creating a loosely applied false capsule for the thyroid. Anterior to the thyroid lobes are the strap muscles. Situated on the posterior surface of the lateral lobes of the thyroid gland are the parathyroid glands and the recurrent laryngeal nerves; the latter usually lie in a cleft between the trachea and the esophagus. The lateral lobes of the thyroid are joined by the isthmus that crosses the trachea. A pyramidal lobe is often present. The pyramidal lobe is a long, narrow projection of thyroid tissue extending upward from the isthmus lying on the surface of the thyroid cartilage. It represents a vestige of the embryonic thyroglossal duct.

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Thyroid Vascular Supply

The thyroid has an abundant blood supply. Its four major arteries are the paired superior thyroid arteries, which arise from the external carotid arteries and

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descend several centimeters in the neck to reach the upper poles of each thyroid lobe, where they branch, and the paired inferior thyroid arteries, each of which arises from the thyrocervical trunk of the subclavian artery, runs medially behind the carotid artery and enters the lower or midpart of the thyroid lobe from behind.

5 A fifth artery, the thyroidea ima, is sometimes present; it arises from the arch of the aorta and enters the thyroid in the midline.

A venous plexus forms under the thyroid capsule. Each lobe is drained by the superior thyroid vein at the upper pole and the middle thyroid vein at the middle
10 part of the lobe, both of which enter the internal jugular vein. Arising from each lower pole are the inferior thyroid veins, which drain directly into the innominate vein.

Thyroid Innervation

15 Significantly, the thyroid gland receives innervation from both the sympathetic and parasympathetic divisions of the autonomic nervous system. The sympathetic fibers arise from the cervical ganglia and enter with blood vessels, whereas the parasympathetic fibers are derived from the vagus and reach the gland via branches of the laryngeal nerves. The thyroid gland's relation to the
20 recurrent laryngeal nerves and to the external branch of the superior laryngeal nerves is of major surgical significance, since damage to these nerves can lead to a disability of phonation.

Sympathetic innervation of the thyroid cells has been reported to exert a
25 stimulatory effect upon thyroid hormone release through adrenergic receptors for norepinephrine on follicle cells. *Endocrinology* 1979;105:7-9. Significantly, the human thyroid is also innervated by cholinergic, parasympathetic fibers. *Cell Tiss Res* 1978;195:367-370. See also *Biol Signals* 1994;3:15-25. And other mammalian species are known to also have cholinergically innervated thyroid
30 cells. See e.g. *Z. Mikrosk Anat Forsch Leipzig* 1986;100:1,S, 34-38 (pig thyroid

is cholinergically innervated); *Neuroendocrinology* 1991;53:69-74 (rat thyroid is cholinergically innervated); *Endocrinology* 1984;114:1266-1271 (dog thyroid is cholinergically innervated);

5 It has been reported that stimulation of the vagal nerve increases both thyroid blood flow and thyroid hormone secretion (*Cell Tiss Res* 1978;195:367-370), but this is apparently due to the extensive, generalized effect of vagal stimulation which can trigger a number of reflexes ascribed to the whole vagus territory. It is therefore inappropriate to conclude from this observation the vagal stimulation
10 acts directly upon the thyroid to increase thyroid hormone release.

Significantly, the consensus is that cholinergic, parasympathetic influence upon thyroid hormone secretion by thyroid follicle cells, and presumably also of the intimately associated C cells, is inhibitory. *Endocrinology* 1979;105:7-9;
15 *Endocrinology* 1984;114:1266-1271; *Peptides* 1985;6:585-589; *Peptides* 1987;8:893-897, and; *Brazilian J Med Biol Res* 1994;27:573-599. The direct cholinergic influence upon the thyroid appears to be mediated by muscarinic acetylcholine receptors of thyroid follicle cells since the cholinergic inhibition is blocked by atropine. *Endocrinology* 1979;105:7. The proximity of the non-
20 follicular, calcitonin secreting cells of the thyroid to the thyroid hormone secreting follicle cells has led to the conclusion that parasympathetic influence over the C cells is also inhibitory.

Current Therapy

25 Therapy for thyroid disorders includes systemically administered drugs, radiotherapy and surgical resection. Unfortunately, all three of these current therapeutic procedures to treat thyroid disorders, including Graves' disease, have significant drawbacks and deficiencies.

Drug therapy for hyperthyroidism includes use of the antithyroid drugs propylthiouracil (PTU) and methimazole (Tapazole), both of which inhibit the organic binding of iodide. In addition, propylthiouracil inhibits the peripheral conversion of T_4 to T_3 . Notably, the half-life of propylthiouracil is only about 1.5 hours while that of methimazole is only about 6 hours. The initial dose of propylthiouracil is 200 to 300 mg, up to 1200 mg daily, every 8 to 12 hours or every 4 to 6 hours when large doses are required. The usual regimen of methimazole is 20 to 40 mg daily in one to three divided doses. Such drug therapy is usually prescribed for 12 to 18 months.

As is well known, intravenous, oral or other systemic routes for drug administration can cause many undesirable side effects, including nausea, diarrhea and drug resistance. Additionally, the patient must remember to take the medication. While the antithyroid drugs for treating hyperthyroidism, such as carbimazole, methimazole and propylthiouracil, suppress the ability of the thyroid to make hormones and can render the patient euthyroid, they can also cause nausea, indigestion, skin rashes, joint pain, fever, and lymphatic gland swelling. Additional side effects include neutropenia and agranulocytosis. Beta adrenergic blocking drugs, such as propranolol (taken orally several times a day), have been used to treat secondary symptoms of hyperthyroidism, such as excessive sweating, tachycardia, hand tremors and anxiety. Iodine has also been used to treat hyperthyroidism due to its suppressive effect upon the release of thyroid hormones. Unfortunately, this effect lasts for only about 3 or 4 weeks.

Radioactive iodine therapy using ^{131}I therapy is designed to administer a sufficient radiation dose to partially destroy the thyroid parenchyma. Biologic effects of ^{131}I include pyknosis and necrosis of the follicular cells and, later, vascular and stromal fibrosis. The ^{131}I dose, in microcuries (μCi), to deliver is calculated using the formula: weight of thyroid gland (gm) X dose

($\mu\text{Ci/gm}$)/uptake (%). The weight of the thyroid gland is estimated by palpation, the 24 hour iodine uptake is measured using a tracer dose of ^{123}I . The dose of ^{131}I that is used for treatment of Graves' disease ranges from 70 to 215 $\mu\text{Ci/gm}$. Higher doses are associated with less relapse but can also be associated with a higher incidence of hypothyroidism during the first few years following treatment.

Radiation therapy, such as by use of radioactive iodine (^{131}I) which concentrates in the thyroid, can by irradiation destroy healthy tissue and cause toxicity reactions, and radiation therapy can have potential harmful effects.

Notably, iodine therapy is not used beyond the first trimester of pregnancy or in children under 15. Additionally, it can take 2 or 3 months before an effect of radio-iodine treatment becomes apparent. Furthermore, the destruction of thyroid cells by result in later hypothyroidism.

Significantly, about 80 percent of Graves' disease patients treated with ^{131}I subsequently become hypothyroid. Additionally, there is evidence that the treatment of thyroid overactivity in Graves' disease with radio-iodine may aggravate the various ophthalmopathic conditions more than does treatment with an antithyroid drug. Bayliss et al., *supra* pages 69-70.

The usual surgical treatment of Graves disease consists of sub-total thyroidectomy leaving 3 to 5 grams of residual thyroid tissue attached to an intact inferior thyroid artery. The choice of therapy may be influenced by cost, age, the size of the goiter, the degree of thyrotoxicosis, pregnancy status, patient preferences, and response to initial treatment. Surgery, because of the potential complications and the cosmetic effect has only a minimal role in the treatment of Graves' disease and is recommended only in patients for whom other therapies are contraindicated or refused.

The surgical option, that is thyroidectomy, has been performed as therapy for thyrotoxicosis, to remove benign and malignant tumors, to alleviate pressure symptoms or respiratory obstruction attributable to the thyroid, and occasionally, to remove an unsightly goiter. Video assisted thyroidectomy has been used to minimize the length of incisions in the neck or to hide the incisions by placing them below the clavicle or far lateral in the neck. The recurrent laryngeal nerve and parathyroid glands can be seen as the thyroid lobectomy is performed. While surgical removal of dysfunctional thyroid tissue can be an effective therapy, it is irreversible and depends to a large extent upon the skill of the surgeon.

Additionally, some thyroid tumors and cancers are inoperable due to proximity or attachment to vital structures. Furthermore, although the mortality rate accompanying thyroidectomy is very low (reported as 0.19 percent), the morbidity rate is about 13 percent when all complications, including the most minor types are considered.

Complications ensuing from surgery have included hypothyroidism, thyroid storm, wound infection, wound hemorrhage with hematoma formation, recurrent laryngeal nerve injury, hypoparathyroidism and tracheomalacia. Significantly, following total or near-total thyroidectomy patients must take thyroid hormone replacement for the remainder of their lives or suffer severe symptoms and signs of myxedema, including tiredness, weakness, depression, psychosis, mental retardation, coma and death.

Notably, about 20% of hyperthyroid patients who have had a surgical thyroidectomy become hypothyroid within one year. Additionally, damage to the recurrent laryngeal nerves during surgery can cause permanent hoarseness of the voice. Furthermore, intraoperative damage to the parathyroids can cause blood calcium levels to fall with resulting tetany.

Thyroid underactivity can be due to a dietary lack of iodine, the lack of which prevents thyroid cell synthesis of the thyroid hormones. In the Western world, Hashimoto's disease is the most common cause of hypothyroidism.

Hypothyroidism can also result from radio-iodine treatment or surgery to correct thyroid overactivity, as well as to a pituitary disorder. The treatment of choice for hypothyroidism is replacement therapy with thyroxine. Treatment of hypothyroidism by thyroid hormone replacement requires long term, daily dosing with expensive medication from which undesirable side effects can occur.

Ophthalmopathy

Hyperthyroidism can result in an overactivity of the sympathetic nervous system with resulting eyelid retraction. Other ophthalmic disorders can be the result of autoimmune hyperthyroidism (i.e. Graves' disease and Hashimoto's thyroiditis) when the autoantibodies also affect ocular muscles. Thus, proptosis or exophthalmos (eyes pushed forwards), impaired eye fluid drainage which can cause increased fluid pressure and blindness, ophthalmoplegia (impaired eye muscle control), diplopia (double vision), and blindness. Therapeutic approaches have included beta blockers to reduce lid retraction, surgery to lower the eyelids and to correct the diplopia, corticosteroids (such as prednisone and methylprednisolone) to reduce eye protrusion by suppressing the ocular autoimmune reaction, and X-raying the orbits and surgery to increase the size of the orbits.

Significantly, and as indicated, treatment of hyperthyroidism with ^{131}I can exacerbate ophthalmopathy more than does treatment with an antithyroid drug. Hence, an effective drug treatment can be preferred to use of radio-iodine or surgery with all its potential complications and required skill level.

Thus, each of the drug, radioactive iodine and surgical therapies for treating thyroid disorders has significant attendant risks, complications, drawbacks and

deficiencies. Presently available drugs have only an antithyroid, as opposed to a prothyroid effect, and are administered systemically. Clearly, there is a considerable need for an effective antithyroid drug to treat hyperthyroidism and for a suitable alternative to thyroid hormone replacement to treat hypothyroidism.

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Botulinum Toxin

The anaerobic, gram positive bacterium *Clostridium botulinum* produces a potent polypeptide neurotoxin, botulinum toxin, which causes a neuromuscular illness in humans and animals referred to as botulism. The spores of *Clostridium botulinum* are found in soil and can grow in improperly sterilized and sealed food containers of home based canneries, which are the cause of many of the cases of botulism. The effects of botulism typically appear 18 to 36 hours after eating the foodstuffs infected with a *Clostridium botulinum* culture or spores. The botulinum toxin can apparently pass unattenuated through the lining of the gut and attack peripheral motor neurons. Symptoms of botulinum toxin intoxication can progress from difficulty walking, swallowing, and speaking to paralysis of the respiratory muscles and death.

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Botulinum toxin type A is the most lethal natural biological agent known to man. About 50 picograms of a commercially available botulinum toxin type A (purified neurotoxin complex)¹ is a LD₅₀ in mice (i.e. 1 unit). Interestingly, on a molar basis, botulinum toxin type A is about 1.8 billion times more lethal than diphtheria, about 600 million times more lethal than sodium cyanide, about 30 million times more lethal than cobra toxin and about 12 million times more lethal than cholera. Singh, *Critical Aspects of Bacterial Protein Toxins*, pages 63-84 (chapter 4) of *Natural Toxins II*, edited by B.R. Singh et al., Plenum Press, New York (1976) (where the stated LD₅₀ of botulinum toxin type A of 0.3 ng equals 1 U is corrected for the fact that about 0.05 ng of BOTOX® equals 1 unit). One unit

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¹Available from Allergan, Inc., of Irvine, California under the tradename BOTOX® in 100 unit vials. One unit of BOTOX® contains about 50 picograms of botulinum toxin type A complex.

(U) of botulinum toxin is defined as the LD₅₀ upon intraperitoneal injection into female Swiss Webster mice weighing 18 to 20 grams each.

Seven immunologically distinct botulinum neurotoxins have been characterized, these being respectively botulinum neurotoxin serotypes A, B, C₁, D, E, F and G each of which is distinguished by neutralization with type-specific antibodies. The different serotypes of botulinum toxin vary in the animal species that they affect and in the severity and duration of the paralysis they evoke. For example, it has been determined that botulinum toxin type A is 500 times more potent, as measured by the rate of paralysis produced in the rat, than is botulinum toxin type B. Additionally, botulinum toxin type B has been determined to be non-toxic in primates at a dose of 480 U/kg which is about 12 times the primate LD₅₀ for botulinum toxin type A. Botulinum toxin apparently binds with high affinity to cholinergic motor neurons, is translocated into the neuron and blocks the release of acetylcholine.

Botulinum toxins have been used in clinical settings for the treatment of neuromuscular disorders characterized by hyperactive skeletal muscles. Botulinum toxin type A has been approved by the U.S. Food and Drug Administration for the treatment of blepharospasm, strabismus and hemifacial spasm. Non-type A botulinum toxin serotypes apparently have a lower potency and/or a shorter duration of activity as compared to botulinum toxin type A. Clinical effects of peripheral intramuscular botulinum toxin type A are usually seen within one week of injection. The typical duration of symptomatic relief from a single intramuscular injection of botulinum toxin type A averages about three months.

Although all the botulinum toxins serotypes apparently inhibit release of the neurotransmitter acetylcholine at the neuromuscular junction, they do so by affecting different neurosecretory proteins and/or cleaving these proteins at

different sites. For example, botulinum types A and E both cleave the 25
kiloDalton (kD) synaptosomal associated protein (SNAP-25), but they target
different amino acid sequences within this protein. Botulinum toxin types B, D, F
and G act on vesicle-associated protein (VAMP, also called synaptobrevin), with
5 each serotype cleaving the protein at a different site. Finally, botulinum toxin
type C₁ has been shown to cleave both syntaxin and SNAP-25. These
differences in mechanism of action may affect the relative potency and/or
duration of action of the various botulinum toxin serotypes. Significantly, it is
known that the cytosol of pancreatic islet B cells contains at least SNAP-25
10 (*Biochem J* 1;339 (pt 1): 159-65 (April 1999)), and synaptobrevin (*Mov Disord*
1995 May; 10(3): 376).

The molecular weight of the botulinum toxin protein molecule, for all seven of
the known botulinum toxin serotypes, is about 150 kD. Interestingly, the
15 botulinum toxins are released by Clostridial bacterium as complexes comprising
the 150 kD botulinum toxin protein molecule along with associated non-toxin
proteins. Thus, the botulinum toxin type A complex can be produced by
Clostridial bacterium as 900 kD, 500 kD and 300 kD forms. Botulinum toxin
types B and C₁ is apparently produced as only a 500 kD complex. Botulinum
20 toxin type D is produced as both 300 kD and 500 kD complexes. Finally,
botulinum toxin types E and F are produced as only approximately 300 kD
complexes. The complexes (i.e. molecular weight greater than about 150 kD)
are believed to contain a non-toxin hemagglutinin protein and a non-toxin and
non-toxic nonhemagglutinin protein. These two non-toxin proteins (which along
25 with the botulinum toxin molecule comprise the relevant neurotoxin complex)
may act to provide stability against denaturation to the botulinum toxin molecule
and protection against digestive acids when toxin is ingested. Additionally, it is
possible that the larger (greater than about 150 kD molecular weight) botulinum
toxin complexes may result in a slower rate of diffusion of the botulinum toxin
30 away from a site of intramuscular injection of a botulinum toxin complex.

In vitro studies have indicated that botulinum toxin inhibits potassium cation induced release of both acetylcholine and norepinephrine from primary cell cultures of brainstem tissue. Additionally, it has been reported that botulinum toxin inhibits the evoked release of both glycine and glutamate in primary cultures of spinal cord neurons and that in brain synaptosome preparations botulinum toxin inhibits the release of each of the neurotransmitters acetylcholine, dopamine, norepinephrine, CGRP and glutamate.

Botulinum toxin type A can be obtained by establishing and growing cultures of *Clostridium botulinum* in a fermenter and then harvesting and purifying the fermented mixture in accordance with known procedures. All the botulinum toxin serotypes are initially synthesized as inactive single chain proteins which must be cleaved or nicked by proteases to become neuroactive. The bacterial strains that make botulinum toxin serotypes A and G possess endogenous proteases and serotypes A and G can therefore be recovered from bacterial cultures in predominantly their active form. In contrast, botulinum toxin serotypes C₁, D and E are synthesized by nonproteolytic strains and are therefore typically unactivated when recovered from culture. Serotypes B and F are produced by both proteolytic and nonproteolytic strains and therefore can be recovered in either the active or inactive form. However, even the proteolytic strains that produce, for example, the botulinum toxin type B serotype only cleave a portion of the toxin produced. The exact proportion of nicked to unnicked molecules depends on the length of incubation and the temperature of the culture. Therefore, a certain percentage of any preparation of, for example, the botulinum toxin type B toxin is likely to be inactive, possibly accounting for the known significantly lower potency of botulinum toxin type B as compared to botulinum toxin type A. The presence of inactive botulinum toxin molecules in a clinical preparation will contribute to the overall protein load of the preparation, which has been linked to increased antigenicity, without contributing to its clinical

efficacy. Additionally, it is known that botulinum toxin type B has, upon intramuscular injection, a shorter duration of activity and is also less potent than botulinum toxin type A at the same dose level.

5 High quality crystalline botulinum toxin type A can be produced from the Hall A strain of *Clostridium botulinum* with characteristics of $\geq 3 \times 10^7$ U/mg, an A_{260}/A_{278} of less than 0.60 and a distinct pattern of banding on gel electrophoresis. The known Shantz process can be used to obtain crystalline botulinum toxin type A, as set forth in Shantz, E.J., et al, *Properties and use of*
10 *Botulinum toxin and Other Microbial Neurotoxins in Medicine*, Microbiol Rev. 56: 80-99 (1992). Generally, the botulinum toxin type A complex can be isolated and purified from an anaerobic fermentation by cultivating *Clostridium botulinum* type A in a suitable medium. The known process can also be used, upon separation out of the non-toxin proteins, to obtain pure botulinum toxins, such as
15 for example: purified botulinum toxin type A with an approximately 150 kD molecular weight with a specific potency of $1-2 \times 10^8$ LD₅₀ U/mg or greater; purified botulinum toxin type B with an approximately 156 kD molecular weight with a specific potency of $1-2 \times 10^8$ LD₅₀ U/mg or greater, and; purified botulinum toxin type F with an approximately 155 kD molecular weight with a specific
20 potency of $1-2 \times 10^7$ LD₅₀ U/mg or greater.

Already prepared and purified botulinum toxins and toxin complexes can be obtained from List Biological Laboratories, Inc., Campbell, California; the Centre for Applied Microbiology and Research, Porton Down , U.K.; Wako (Osaka,
25 Japan), as well as from Sigma Chemicals of St Louis, Missouri.

Pure botulinum toxin is so labile that it is generally not used to prepare a pharmaceutical composition. Furthermore. the botulinum toxin complexes, such the toxin type A complex are also extremely susceptible to denaturation due to
30 surface denaturation, heat, and alkaline conditions. Inactivated toxin forms

toxoid proteins which may be immunogenic. The resulting antibodies can render a patient refractory to toxin injection.

As with enzymes generally, the biological activities of the botulinum toxins (which are intracellular peptidases) is dependant, at least in part, upon their three dimensional conformation. Thus, botulinum toxin type A is detoxified by heat, various chemicals surface stretching and surface drying. Additionally, it is known that dilution of the toxin complex obtained by the known culturing, fermentation and purification to the much, much lower toxin concentrations used for pharmaceutical composition formulation results in rapid detoxification of the toxin unless a suitable stabilizing agent is present. Dilution of the toxin from milligram quantities to a solution containing nanograms per milliliter presents significant difficulties because of the rapid loss of specific toxicity upon such great dilution. Since the toxin may be used months or years after the toxin containing pharmaceutical composition is formulated, the toxin must be formulated with a stabilizing agent, such as albumin.

A commercially available botulinum toxin containing pharmaceutical composition is sold under the trademark BOTOX® (available from Allergan, Inc., of Irvine, California). BOTOX® consists of a purified botulinum toxin type A complex, albumin and sodium chloride packaged in sterile, vacuum-dried form. The botulinum toxin type A is made from a culture of the Hall strain of Clostridium botulinum grown in a medium containing N-Z amine and yeast extract. The botulinum toxin type A complex is purified from the culture solution by a series of acid precipitations to a crystalline complex consisting of the active high molecular weight toxin protein and an associated hemagglutinin protein. The crystalline complex is re-dissolved in a solution containing saline and albumin and sterile filtered (0.2 microns) prior to vacuum-drying. BOTOX® can be reconstituted with sterile, non-preserved saline prior to intramuscular injection. Each vial of BOTOX® contains about 100 units (U) of Clostridium

botulinum toxin type A purified neurotoxin complex, 0.5 milligrams of human serum albumin and 0.9 milligrams of sodium chloride in a sterile, vacuum-dried form without a preservative.

5 BOTOX® can be reconstituted with 0.9% Sodium Chloride Injection. Since BOTOX® can be denatured by bubbling or similar violent agitation, the diluent is gently injected into the vial. BOTOX® should be administered within four hours after reconstitution. During this time period, reconstituted BOTOX® is stored in a refrigerator (2° to 8°C). Reconstituted BOTOX® is clear, colorless and free of
10 particulate matter. The vacuum-dried product is stored in a freezer at or below -5°C. BOTOX® is administered within four hours after the vial is removed from the freezer and reconstituted. During these four hours, reconstituted BOTOX® can be stored in a refrigerator (2° to 8°C). Reconstituted BOTOX® is clear, colorless and free of particulate matter.

15 It has been reported that botulinum toxin type A has been used in clinical settings as follows:

(1) about 75-125 units of BOTOX® per intramuscular injection (multiple muscles) to treat cervical dystonia;

20 (2) 5-10 units of BOTOX® per intramuscular injection to treat glabellar lines (brow furrows) (5 units injected intramuscularly into the procerus muscle and 10 units injected intramuscularly into each corrugator supercilii muscle);

(3) about 30-80 units of BOTOX® to treat constipation by intrasphincter injection of the puborectalis muscle;

25 (4) about 1-5 units per muscle of intramuscularly injected BOTOX® to treat blepharospasm by injecting the lateral pre-tarsal orbicularis oculi muscle of the upper lid and the lateral pre-tarsal orbicularis oculi of the lower lid.

(5) to treat strabismus, extraocular muscles have been injected intramuscularly with between about 1-5 units of BOTOX®, the amount injected

varying based upon both the size of the muscle to be injected and the extent of muscle paralysis desired (i.e. amount of diopter correction desired).

(6) to treat upper limb spasticity following stroke by intramuscular injections of BOTOX® into five different upper limb flexor muscles, as follows:

(a) flexor digitorum profundus: 7.5 U to 30 U

(b) flexor digitorum sublimus: 7.5 U to 30 U

(c) flexor carpi ulnaris: 10 U to 40 U

(d) flexor carpi radialis: 15 U to 60 U

(e) biceps brachii: 50 U to 200 U. Each of the five indicated muscles has

been injected at the same treatment session, so that the patient receives from 90 U to 360 U of upper limb flexor muscle BOTOX® by intramuscular injection at each treatment session.

(7) to treat migraine, pericranial injected (injected symmetrically into glabellar, frontalis and temporalis muscles) injection of 25 U of BOTOX® has showed significant benefit as a prophylactic treatment of migraine compared to vehicle as measured by decreased measures of migraine frequency, maximal severity, associated vomiting and acute medication use over the three month period following the 25 U injection.

The success of botulinum toxin type A to treat a variety of clinical conditions has led to interest in other botulinum toxin serotypes. A study of two commercially available botulinum type A preparations (BOTOX® and Dysport®) and preparations of botulinum toxins type B and F (both obtained from Wako Chemicals, Japan) has been carried out to determine local muscle weakening efficacy, safety and antigenic potential. Botulinum toxin preparations were injected into the head of the right gastrocnemius muscle (0.5 to 200.0 units/kg) and muscle weakness was assessed using the mouse digit abduction scoring assay (DAS). ED₅₀ values were calculated from dose response curves.

Additional mice were given intramuscular injections to determine LD₅₀ doses.

The therapeutic index was calculated as LD₅₀/ED₅₀. Separate groups of mice

received hind limb injections of BOTOX® (5.0 to 10.0 units/kg) or botulinum toxin type B (50.0 to 400.0 units/kg), and were tested for muscle weakness and increased water consumption, the later being a putative model for dry mouth.

Antigenic potential was assessed by monthly intramuscular injections in rabbits

(1.5 or 6.5 ng/kg for botulinum toxin type B or 0.15 ng/kg for BOTOX®). Peak muscle weakness and duration were dose related for all serotypes. DAS ED₅₀ values (units/kg) were as follows: BOTOX®: 6.7, Dysport®: 24.7, botulinum toxin type B: 27.0 to 244.0, botulinum toxin type F: 4.3. BOTOX® had a longer duration of action than botulinum toxin type B or botulinum toxin type F.

Therapeutic index values were as follows: BOTOX®: 10.5, Dysport®: 6.3, botulinum toxin type B: 3.2. Water consumption was greater in mice injected with botulinum toxin type B than with BOTOX®, although botulinum toxin type B was less effective at weakening muscles. After four months of injections 2 of 4 (where treated with 1.5 ng/kg) and 4 of 4 (where treated with 6.5 ng/kg) rabbits developed antibodies against botulinum toxin type B. In a separate study, 0 of 9 BOTOX® treated rabbits demonstrated antibodies against botulinum toxin type A. DAS results indicate relative peak potencies of botulinum toxin type A being equal to botulinum toxin type F, and botulinum toxin type F being greater than botulinum toxin type B. With regard to duration of effect, botulinum toxin type A was greater than botulinum toxin type B, and botulinum toxin type B duration of effect was greater than botulinum toxin type F. As shown by the therapeutic index values, the two commercial preparations of botulinum toxin type A (BOTOX® and Dysport®) are different. The increased water consumption behavior observed following hind limb injection of botulinum toxin type B indicates that clinically significant amounts of this serotype entered the murine systemic circulation. The results also indicate that in order to achieve efficacy comparable to botulinum toxin type A, it is necessary to increase doses of the other serotypes examined. Increased dosage can compromise safety.

Furthermore, in rabbits, type B was more antigenic than was BOTOX®, possibly

because of the higher protein load injected to achieve an effective dose of botulinum toxin type B. *Eur J Neurol* 1999 Nov;6(Suppl 4):S3-S10.

Acetylcholine

Typically only a single type of small molecule neurotransmitter is released by each type of neuron in the mammalian nervous system. The neurotransmitter acetylcholine is secreted by neurons in many areas of the brain, but specifically by the large pyramidal cells of the motor cortex, by several different neurons in the basal ganglia, by the motor neurons that innervate the skeletal muscles, by the preganglionic neurons of the autonomic nervous system (both sympathetic and parasympathetic), by the postganglionic neurons of the parasympathetic nervous system, and by some of the postganglionic neurons of the sympathetic nervous system. Essentially, only the postganglionic sympathetic nerve fibers to the sweat glands, the piloerector muscles and a few blood vessels are cholinergic as most of the postganglionic neurons of the sympathetic nervous system secrete the neurotransmitter norepinephrine. In most instances acetylcholine has an excitatory effect. However, acetylcholine is known to have inhibitory effects at some of the peripheral parasympathetic nerve endings, such as inhibition of heart rate by the vagal nerve.

The efferent signals of the autonomic nervous system are transmitted to the body through either the sympathetic nervous system or the parasympathetic nervous system. The preganglionic neurons of the sympathetic nervous system extend from preganglionic sympathetic neuron cell bodies located in the intermediolateral horn of the spinal cord. The preganglionic sympathetic nerve fibers, extending from the cell body, synapse with postganglionic neurons located in either a paravertebral sympathetic ganglion or in a prevertebral ganglion. Since, the preganglionic neurons of both the sympathetic and parasympathetic nervous system are cholinergic, application of acetylcholine to

the ganglia will excite both sympathetic and parasympathetic postganglionic neurons.

Acetylcholine activates two types of receptors, muscarinic and nicotinic receptors. The muscarinic receptors are found in all effector cells stimulated by the postganglionic neurons of the parasympathetic nervous system, as well as in those stimulated by the postganglionic cholinergic neurons of the sympathetic nervous system. The nicotinic receptors are found in the synapses between the preganglionic and postganglionic neurons of both the sympathetic and parasympathetic. The nicotinic receptors are also present in many membranes of skeletal muscle fibers at the neuromuscular junction.

Acetylcholine is released from cholinergic neurons when small, clear, intracellular vesicles fuse with the presynaptic neuronal cell membrane. A wide variety of non-neuronal secretory cells, such as, adrenal medulla (as well as the PC12 cell line) and pancreatic islet cells release catecholamines and thyroid hormone, respectively, from large dense-core vesicles. The PC12 cell line is a clone of rat pheochromocytoma cells extensively used as a tissue culture model for studies of sympathoadrenal development. Botulinum toxin inhibits the release of both types of compounds from both types of cells *in vitro*, permeabilized (as by electroporation) or by direct injection of the toxin into the denervated cell. Botulinum toxin is also known to block release of the neurotransmitter glutamate from cortical synaptosomes cell cultures.

What is needed therefore is an effective, long lasting, non-surgical resection, non-radiotherapy, non-systemic drug administration, therapeutic drug and method for treating thyroid disorders.

SUMMARY

The present invention meets this need and provides an effective, non-surgical resection, relatively long term, non-radiotherapy, non-systemic drug administration, therapeutic method for treating thyroid disorders. The drug within the scope of this invention for treating thyroid disorders is a neurotoxin. Significantly, the same neurotoxin can be used to treat hypothyroidism, hyperthyroidism, hypocalcemia and hypercalcemia depending upon factors such as the site of local administration of the neurotoxin and the amount of neurotoxin to be administered.

As used herein "local administration" means direct injection of a neurotoxin into the thyroid or into a sympathetic ganglion which innervates a thyroid cell (such as a thyroid follicle cell or thyroid C cell). Systemic routes of administration, such as oral and intravenous routes of administration, are excluded from the scope of "local administration" of a neurotoxin.

As used herein, "thyroid hormone" means thyroxine (T_4), while "thyroid hormones" means triiodothyronine (T_3) and thyroxine (T_4)

A method for treating a thyroid disorder according to the present invention can be carried out by administration of a therapeutically effective amount of a neurotoxin to a patient, thereby treating the thyroid disorder. The neurotoxin can administered to the thyroid of the patient when the thyroid disorder to be treated is hypothyroidism. Alternately, the neurotoxin can be administered to a sympathetic ganglion which innervates the thyroid when the thyroid disorder to be treated is hyperthyroidism.

A detailed method for treating a thyroid disorder according to the present invention can comprise the step of administration of a therapeutically effective

amount of a botulinum toxin to a patient. Thus, a method for treating hypothyroidism according to the present invention can comprise the step of local administration to the thyroid of a therapeutically effective amount of a botulinum toxin, thereby increasing a deficient thyroid hormone secretion from the thyroid cell and effectively treating the hypothyroidism. Furthermore, a method within the scope of the present invention for treating hyperthyroidism, can comprise the step of local administration to a sympathetic ganglion which innervates a thyroid cell of a therapeutically effective amount of a botulinum toxin, thereby reducing an excessive thyroid hormone secretion from the thyroid cell and hence effectively treating the hyperthyroidism.

The neurotoxin can be administered in an amount of between about 10^{-3} U/kg and about 35 U/kg. 35 U/kg is an upper limit because it approaches a lethal dose of certain neurotoxins, such as botulinum toxin type A. Other botulinum toxins, such as botulinum toxin type B, can be safely administered at several orders of magnitude higher dosage. Preferably, the neurotoxin is administered in an amount of between about 10^{-2} U/kg and about 25 U/kg. More preferably, the neurotoxin is administered in an amount of between about 10^{-1} U/kg and about 15 U/kg. Most preferably, the neurotoxin is administered in an amount of between about 1 U/kg and about 10 U/kg. In many instances, an administration of from about 1 units to about 500 units of a neurotoxin, such as a botulinum toxin type A, provides effective and long lasting therapeutic relief. More preferably, from about 5 units to about 300 units of a neurotoxin, such as a botulinum toxin type A, can be used and most preferably, from about 10 units to about 200 units of a neurotoxin, such as a botulinum toxin type A, can be locally administered into a target tissue such as the thyroid or a sympathetic ganglion with efficacious results. In a particularly preferred embodiment of the present invention from about 10 units to about 100 units of a botulinum toxin, such as botulinum toxin type A, can be locally administered into a target tissue such as the thyroid or a sympathetic ganglion with therapeutically effective results.

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The neurotoxin can be made by a Clostridial bacterium, such as by a *Clostridium botulinum*, *Clostridium butyricum*, *Clostridium beratti* or *Clostridium tetani* bacterium. Additionally, the neurotoxin can be a modified neurotoxin, that is a neurotoxin that has at least one of its amino acids deleted, modified or replaced, as compared to the native or wild type neurotoxin. Furthermore, the neurotoxin can be a recombinant produced neurotoxin or a derivative or fragment thereof.

The neurotoxin can be a botulinum toxin, such as one of the botulinum toxin serotypes A, B, C₁, D, E, F or G. Preferably, the neurotoxin is botulinum toxin type A and the neurotoxin is locally administered by direct injection of the neurotoxin into the thyroid or into a sympathetic ganglion which innervates the thyroid.

A detailed embodiment of a method within the scope of the present invention for treating a thyroid disorder can comprise the step of injecting a therapeutically effective amount of a botulinum toxin into a thyroid of a human patient, thereby increasing a thyroid hormone secretion from a thyroid cell and treating a thyroid disorder.

Another detailed embodiment of a method within the scope of the present invention for treating a thyroid disorder of a human patient can comprise the step of local administration to a cholinergic influenced thyroid cell of a human patient of a therapeutically effective amount of botulinum toxin type A, thereby increasing a cholinergic influenced deficient thyroid hormone secretion from the thyroid cell and treating the thyroid disorder.

Another method within the scope of the present invention is a method for treating a thyroid disorder by administration of a neurotoxin to a sympathetic

nervous system of a patient. In this method the neurotoxin is locally administered to a sympathetic ganglion which innervates a thyroid cell and the thyroid disorder is hyperthyroidism.

5 A detailed embodiment of a method within the scope of the present invention for treating a thyroid disorder of a human patient can comprise the step of *in vivo*, local administration to a sympathetic ganglion, which innervates a thyroid cell of a patient, of a therapeutically effective amount of a botulinum toxin, thereby decreasing an excessive thyroid hormone secretion from a thyroid cell
10 and treating hyperthyroidism.

A detailed embodiment of the present invention is a method for treating a thyroid disorder by injecting a therapeutically effective amount of a botulinum toxin into a thyroid of a human patient, thereby increasing a secretion of a thyroid
15 hormone from a thyroid cell and treating the thyroid disorder. Preferably, the secretion treated is a cholinergic influenced secretion and the botulinum toxin used is botulinum toxin type A, although the botulinum toxin can be selected from the group consisting of botulinum toxin types A, B, C (i.e. C₁), D, E, F and G.

20 My invention also includes within its scope, a method for treating hypercalcemia, the method comprising the step of local administration to a thyroid C cell of a therapeutically effective amount of a botulinum toxin, thereby increasing a deficient calcitonin secretion from a thyroid C cell and treating hypercalcemia. Additionally, my invention also includes within its scope a
25 method for treating hypocalcemia, the method comprising the step of local administration to a sympathetic ganglion which innervates a thyroid C cell of a therapeutically effective amount of a botulinum toxin, thereby increasing a deficient calcitonin secretion from the thyroid C cell and treating hypocalcemia.

DESCRIPTION

The present invention is based upon the discovery that a thyroid disorder can be treated by *in vivo* administration of a neurotoxin to a patient. Thus

5 administration of a neurotoxin to the thyroid of a patient can remove an inhibitory cholinergic effect upon thyroid hormone secretion, thereby providing an effective treatment for hypothyroidism. Additionally, administration of a neurotoxin to a sympathetic ganglion which innervates the thyroid can remove a stimulatory adrenergic effect upon thyroid hormone secretion, thereby providing an effective
10 treatment for hyperthyroidism.

Thus, thyroid disorders can be treated, according to the present invention, by the alternative therapies of (a) local administration of a neurotoxin to the thyroid, or; (b) local administration of a neurotoxin to a sympathetic ganglion of a patient,
15 thereby resulting in, respectively, an increase of a secretion from a thyroid cell, or a decrease in a secretion from a thyroid cell

I have discovered that a particular neurotoxin, botulinum toxin, can be used with dramatic ameliorative effect to treat a thyroid disorder, thereby significantly
20 superseding thereby current therapeutic regimens, such as oral thyroid hormone (to treat hypothyroidism) or radioactive iodine (to treat hyperthyroidism). Significantly, a single local administration of a neurotoxin, such as a botulinum toxin to the thyroid, according to the present invention, can increase thyroid hormone secretion and thereby treat symptoms of hypothyroidism. I have also
25 discovered that a single local administration of a neurotoxin, such as a botulinum toxin to a sympathetic ganglion which innervates the thyroid gland, according to the present invention, can reduce thyroid hormone secretion and thereby treat symptoms of hyperthyroidism. In either case, the symptoms of the thyroid disorder can be alleviated for at least about from 2 months to about 6 months per
30 neurotoxin administration. Notably, it has been reported that glandular tissue

as for example 10 to 500 units of botulinum toxin type A, to cholinergic, postganglionic, parasympathetic neurons which innervate the dysfunctional, thyroid cells. Without wishing to be bound by theory, the botulinum toxin is believed to act by inhibiting release of acetylcholine neurotransmitter from cholinergic, postganglionic parasympathetic fibers which innervate thyroid follicle cells.

A neurotoxin, such as a botulinum toxin, locally administered *in vivo* to the thyroid to thereby remove an inhibitory effect upon a secretory activity of a thyroid follicle cell. The thyroid follicle cell is cholinergically innervated or susceptible to high toxin dosing such that the proteolytic light chain of the toxin is internalized by a cholinergic neuron which influences a secretory activity of the thyroid cell.

Thus, cholinergically innervated thyroid cells can be treated by local administration of a neurotoxin, such as a botulinum toxin. By local administration it is meant that the neurotoxin is administered directly to or to the immediate vicinity of the thyroid tissue to be treated.

The specific dosage appropriate for administration is readily determined by one of ordinary skill in the art according to the factor discussed above. The dosage can also depend upon the size of the thyroid tissue mass to be treated or denervated, and the commercial preparation of the toxin. Additionally, the estimates for appropriate dosages in humans can be extrapolated from determinations of the amounts of botulinum required for effective denervation of other tissues. Thus, the amount of botulinum A to be injected is proportional to the mass and level of activity of the thyroid tissue to be treated. Generally, between about 0.01 and 35 units per kg of patient weight of a botulinum toxin, such as botulinum toxin type A, can be administered to effectively accomplish a toxin induced thyroid tissue secretion up regulation upon administration of the

neurotoxin into the thyroid. Less than about 0.01 U/kg of a botulinum toxin does not have a significant therapeutic effect upon the secretory activity of a thyroid cell, while more than about 35 U/kg of a botulinum toxin approaches a toxic dose the neurotoxin. Careful placement of the injection needle and a low volume of neurotoxin used prevents significant amounts of botulinum toxin from appearing systemically. A more preferred dose range is from about 0.01 U/kg to about 25 U/kg of a botulinum toxin, such as that formulated as BOTOX®. The actual amount of U/kg of a botulinum toxin to be administered depends upon factors such as the extent (mass) and level of activity of the thyroid tissue to be treated and the administration route chosen. Botulinum toxin type A is a preferred botulinum toxin serotype for use in the methods of the present invention.

It has been reported that the neuronal selectivity of clostridial neurotoxins is a result of a very selective binding and cell entry mechanism. Although a site of action of botulinum toxin is the neuromuscular junction, where the toxin binds rapidly and prevents the release of acetylcholine from cholinergic neurons, it is known that clostridial neurotoxins are able to enter certain neurosecretory cells (for example PC12 cells) via a low affinity receptor if high concentrations of the neurotoxin are incubated with the cells for prolonged periods. This process appears to use a pathway via a receptor which is distinct from the highly specific and high affinity receptor present at the neuromuscular junction. Additionally, it has been reported that certain clostridial toxins have effects on phagocyte cells, such as macrophages, where entry into the cell is presumed to be via the specific phagocytic activity of these cells. Furthermore, incubation of certain adipocytes (i.e. fat cells) with botulinum toxin type A has been reported to inhibit glucose uptake by the adipocytes. The mechanism of the glucose uptake inhibition is apparently due to toxin inhibition of plasma membrane fusion or docking of cytosolic, recyclable membrane vesicles (RMVs), the RMVs containing glucose transporter proteins. *PCT publication WO 94/21300.*

Thus, while it is known that the botulinum toxins have a known binding affinity for cholinergic, pre-synaptic, peripheral motor neurons, it has been reported that botulinum toxins can also bind to and translocate into a variety of non-neuronal secretory cells, where the toxin then acts, in the known manner, as an endoprotease upon its respective secretory vessel-membrane docking protein. Because of the relatively lower affinity of the botulinum toxins for secretory cells, such as thyroid cells, as compared to the affinity of the botulinum toxin for the cholinergic neurons which innervate thyroid cells, the botulinum toxin can be injected into secretory or glandular tissues to provide a high local concentration of the toxin, thereby facilitating effect of the toxin upon both cholinergic neuron and directly upon thyroid secretory cell. Thus, the present invention is applicable to the treatment of thyroid disorders wherein the target thyroid cells have little or no cholinergic innervation.

Preferably, a neurotoxin used to practice a method within the scope of the present invention is a botulinum toxin, such as one of the serotype A, B, C, D, E, F or G botulinum toxins. Preferably, the botulinum toxin used is botulinum toxin type A, because of its high potency in humans, ready availability, and known safe and efficacious use for the treatment of skeletal muscle and smooth muscle disorders when locally administered by intramuscular injection.

The present invention includes within its scope the use of any neurotoxin which has a long duration therapeutic effect when locally applied to treat a thyroid cell disorder of a patient. For example, neurotoxins made by any of the species of the toxin producing Clostridium bacteria, such as *Clostridium botulinum*, *Clostridium butyricum*, and *Clostridium beratti* can be used or adapted for use in the methods of the present invention. Additionally, all of the botulinum serotypes A, B, C, D, E, F and G can be advantageously used in the practice of the present invention, although type A is the most preferred serotype, as

explained above. Practice of the present invention can provide effective relief of a thyroid disorder for from 2-27 months or longer in humans.

Botulinum toxin is believed to be able to block the release of any vesicle mediated exocytosis from any secretory (i.e. neuronal, glandular, secretory, chromaffin) cell type, as long as the light chain of the botulinum toxin is translocated into the intracellular medium. For example, the intracellular protein SNAP-25 is widely distributed in both neuronal and non-neuronal secretory cells and botulinum toxin type A is an endopeptidase for which the specific substrate is SNAP-25. Thus, while cholinergic neurons have a high affinity receptor for the botulinum and tetanus toxins (and are therefore more sensitive than other neurons and other cells to the inhibition of vesicle mediated exocytosis of secretory compounds), as the toxin concentration is raised, non-cholinergic sympathetic neurons, chromaffin cells and other cell types can take up a botulinum toxin and show reduced exocytosis.

Hence, by practice of the present disclosed invention, non-cholinergic nerve fibers as well as non or poorly innervated thyroid cells can be treated by use of an appropriately higher concentration of a botulinum toxin to bring about therapeutic relief from a thyroid disorder.

Local Administration of a Neurotoxin to a Sympathetic Ganglion

Significantly, a method within the scope of the present invention for reducing an excessive thyroid hormone secretion comprises the step of local administration of a neurotoxin to the sympathetic nervous system. Sympathetic innervation of the thyroid is known to exist. Thus, sympathetic nerve fibers can inhibit thyroid hormone secretion by acting via adrenergic receptors on thyroid follicle. A method within the scope of the present invention can therefore be carried out by local administration of a neurotoxin to a cholinergic, preganglionic sympathetic neuron. The cholinergic, preganglionic, sympathetic neurons

synapse with adrenergic, post-ganglionic, sympathetic fibers, and these later sympathetic neurons have a stimulatory effect upon thyroid hormone secretion by thyroid cells. Preferably, the sympathetic ganglion to which a neurotoxin is administered, according to the preset invention, is a cervical ganglion.

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Cervical ganglion block according to the present invention can be carried out in the same manner as a celiac plexus block. Thus, the neurolytic celiac plexus block is a known procedure for treating intractable pain resulting from upper abdominal viscera cancer. *Reg Anest Pain Med* 1998; 23(1):37-48. Thus, it is known to inject the celiac plexus with ethanol or phenol to provide relief from the pain which can result from pancreatic cancer or from pancreatitis. *AJG* 1999;94(4):872-874. Hence, an antinociceptive injection of the cervical ganglia can be carried out as by either a percutaneous procedure or as an open (intraoperative) injection. The percutaneous (closed) procedure can be carried out using an anterior approach using a very thin needle (22 Gauge). Cervical ganglion block is preferably carried out with computed tomography (CT) (as opposed to fluoroscopic) needle guidance, using a single thin needle.

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Furthermore, a method within the scope of the present invention can provide improved patient function. "Improved patient function" can be defined as an improvement measured by factors such as a reduced pain, reduced time spent in bed, increased ambulation, healthier attitude, more varied lifestyle and/or healing permitted by normal muscle tone. Improved patient function is synonymous with an improved quality of life (QOL). QOL can be assessed using, for example, the known SF-12 or SF-36 health survey scoring procedures. SF-36 assesses a patient's physical and mental health in the eight domains of physical functioning, role limitations due to physical problems, social functioning, bodily pain, general mental health, role limitations due to emotional problems, vitality, and general health perceptions. Scores obtained can be compared to published values available for various general and patient populations.

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As set forth above, I have discovered that a surprisingly effective and long lasting therapeutic effect can be achieved by local administration of a neurotoxin to the thyroid or to a sympathetic ganglion which innervates the thyroid of a human patient. In its most preferred embodiment, the present invention is practiced by direct injection into the thyroid or into the sympathetic ganglion of botulinum toxin type A. It has been reported that at the neuroglandular junction, the chemical denervation effect of a botulinum toxin, such as botulinum toxin type A, has a long duration of action, i.e. 27 months vs. 3 months.

The present invention includes within its scope: (a) neurotoxin complex as well as pure neurotoxin obtained or processed by bacterial culturing, toxin extraction, concentration, preservation, freeze drying and/or reconstitution and; (b) modified or recombinant neurotoxin, that is neurotoxin that has had one or more amino acids or amino acid sequences deliberately deleted, modified or replaced by known chemical/biochemical amino acid modification procedures or by use of known host cell/recombinant vector recombinant technologies, as well as derivatives or fragments of neurotoxins so made, and includes neurotoxins with one or more attached targeting moieties for a cell surface receptor present on a thyroid cell.

Botulinum toxins for use according to the present invention can be stored in lyophilized or vacuum dried form in containers under vacuum pressure. Prior to lyophilization the botulinum toxin can be combined with pharmaceutically acceptable excipients, stabilizers and/or carriers, such as albumin. The lyophilized or vacuum dried material can be reconstituted with saline or water.

The route of administration and amount of a neurotoxin (such as a botulinum toxin serotype A, B, C, D, E, F or G) administered according to the present invention for treating a thyroid disorder can vary widely according to various

patient variables including size, weight, age, disease severity, responsiveness to therapy, and solubility and diffusion characteristics of the neurotoxin toxin chosen. Furthermore, the extent of the thyroid or ganglionic tissue influenced is believed to be proportional to the volume of neurotoxin injected, while the quantity of the denervation is, for most dose ranges, believed to be proportional to the concentration of neurotoxin injected.

Methods for determining the appropriate route of administration and dosage are generally determined on a case by case basis by the attending physician.

Such determinations are routine to one of ordinary skill in the art (see for example, *Harrison's Principles of Internal Medicine* (1998), edited by Anthony Fauci et al., 14th edition, published by McGraw Hill). For example, to treat a thyroid disorder, a solution of botulinum toxin type A complex can be endoscopically or intraperitoneally injected directly into the tissues of the thyroid, thereby substantially avoiding entry of the toxin into the systemic circulation.

EXAMPLES

The following examples provide those of ordinary skill in the art with specific preferred methods within the scope of the present invention for carrying out the present invention and are not intended to limit the scope of what the inventor regards as his invention. In each of the following examples, the specific amount of a botulinum toxin administered depends upon a variety of factors to be weighed and considered within the discretion of the attending physician and in each of the examples insignificant amounts of botulinum toxin enter appear systemically with no significant side effects. Units of botulinum toxin injected per kilogram (U/kg) below are per kg of total patient weight. For example, 3U/kg for a 70 kg patient calls for an injection of 210 units of the botulinum toxin.

Example 1
Intraoperative Administration of Neurotoxin

Intraoperative, local administration of a neurotoxin to the thyroid can be carried out as follows. The procedure can be performed under general endotracheal anesthesia. The patient's neck can be extended by inflating a pillow or inserting a thyroid roll beneath the shoulders. A symmetrical, low, collar incision can then be made in the line of a natural skin crease approximately 1 to 2 cm above the clavicle. The incision can be carried through the skin, subcutaneous tissue, and platysma muscle down to the dense cervical fascia that overlies the strap muscles and anterior jugular veins. The upper flap can then be raised to a level cephalad to the cricoid cartilage. Care is taken to avoid cutting sensory nerves. A small lower flap is also elevated to the level of the manubrial notch. Performing dissection of the flaps in the plane between the platysma muscle and the fascia overlying the strap muscles results in minimal bleeding. The cervical fascia is then incised vertically in the midline from the thyroid cartilage to the sternal notch.

Exposure of the superior and lateral aspects of the thyroid gland is achieved by retracting the sternohyoid and sternothyroid muscles laterally or in very large glands by dividing these muscles. Division of these muscles is associated with little or no disability, but is not necessary unless the gland is markedly enlarged. High transection is preferable, since the ansa cervicalis nerve innervates the muscles from below. This procedure diminishes the amount of muscle that is paralyzed.

Digital or blunt dissection frees the thyroid from the surrounding fascia. As an initial step, the isthmus of the thyroid is usually revealed. Rotation of one lobe of the thyroid can be followed by dissection bluntly. The middle thyroid veins are first encountered and are ligated and divided. This maneuver facilitates

exposure of the superior and inferior poles of the thyroid lobe. The suspensory ligaments are transected craniad to the isthmus, and the pyramidal lobe and Delphian nodes are mobilized. The cricothyroid space is opened in order to separate the superior pole from the surrounding tissue. If dissection of the superior lobe is to be carried out (i.e. thyroidectomy in conjunction with neurotoxin administration), care is taken to avoid injury to the superior laryngeal nerve. The internal branch of the nerve, which provides sensory fibers to the epiglottis and larynx, is rarely in the operative field. It is the external branch, which supplies motor innervation to the inferior pharyngeal constrictor and the cricothyroid muscle, that must be protected. This purpose is achieved by dissecting the nerve away from the superior pole vessels if it can be identified, or by separately ligating and dividing the individual branches of the superior thyroid vessels immediately adjacent to the upper pole of the thyroid lobe rather than cephalad to it.

The lobe can then be retracted mediad to permit identification of the inferior thyroid artery and the recurrent laryngeal nerve. It is essential that meticulous hemostasis be achieved during this part of the dissection. The inferior thyroid artery is isolated, but need not be ligated laterally. Rather, when performing a lobectomy, it is preferable to ligate and divide each small arterial branch near the thyroid capsule at a point after branches to the parathyroid glands have been given off. This technique lessens the incidence of devascularization of the parathyroid gland and plays a role in reducing permanent hypoparathyroidism. If a parathyroid gland is devascularized it can be minced and autotransplanted into the sternocleidomastoid after being verified by frozen section analysis. The recurrent laryngeal nerve can be identified along its course by blunt dissection. The fibrous tissues are gently unroofed from the front of the nerve. The nerve is treated with care, for excessive trauma or its division will result in an ipsilateral vocal cord paralysis. At the junction of the trachea and larynx, the recurrent laryngeal nerve is immediately adjacent to the thyroid lobe and is in greatest

danger, if it is not seen. The exposed thyroid lobe can now be directed injected with from 10 to 300 units of a botulinum toxin, such as botulinum toxin type A.

During exposure of the posterior surface of the thyroid gland, the parathyroid glands should be identified and preserved, along with their vascular pedicles. Care should be taken to ensure that the parathyroid glands are not excised or devascularized.

The entire wound is then inspected, and careful hemostasis is obtained before closure. In most instances, a suction catheter is used to drain the bed of the thyroid lobes, employing a small, soft plastic drain that is brought out through a stab wound lateral to the incision. The postoperative wound appearance with this technique is far superior to that obtained when no drainage is employed. If the sternothyroid and sternohyoid muscles have been transected, they are reapproximated. The midline vertical fascial incision is only loosely approximated by one interrupted suture, and the drain is positioned superficial to the strap muscles. There is generally no need to suture the platysma muscle, instead it is preferable to approximate the deep dermis with interrupted 4-0 resorbable sutures and the epithelium with 5-0 continuous subcuticular sutures. Finally, the epithelial surfaces are approximated with sterile skin tapes. Within one to seven days, thyroid hormone secretion is substantially increased due to removal of cholinergic inhibition and this effect persists for from 2 to 6 months.

Example 2

Local Administration of Neurotoxin to the Thyroid

Local administration of a neurotoxin directly to or to the vicinity of the thyroid can be accomplished by several methods. For example, by thyroid endoscopy.

An endoscope used for thyroid therapy can be modified to permit its use for direct injection of a neurotoxin, such as a botulinum toxin directly into thyroid

tissue. See for example U.S. patent no. 5,674,205. Once appropriately located, a hollow needle tip can be extended from the endoscope into thyroid tissue and through which needle the neurotoxin can be injected into the thyroid tissue.

Additionally, fine needle aspiration for thyroid biopsy purposes is well known and can be used to inject a neurotoxin, rather than to aspirate thyroid tissue. From 10 to 300 units of a botulinum toxin, such as botulinum toxin type A can thereby be injected into the thyroid. Within one to seven days, thyroid hormone secretion is substantially increased due to removal of cholinergic inhibition and this effect persists for from 2 to 6 months.

Example 3

Treatment of Hypothyroidism With Botulinum Toxin Type A

A 43 year old, obese man presents with increasing fatigue over the last eight months and worsening pedal and calf edema over same time period, which is exacerbated upon standing for very long. Patient also has had polydipsia and low urine output over same time period. Notably, TSH is 690 (normal is 3-5). A diagnosis of hypothyroidism is made. Between about 10^{-3} U/kg and about 35 U/kg of a botulinum toxin type A preparation (for example between about 10 units and about 300 units of BOTOX®) is injected directly into the thyroid, using one of the techniques set forth in Examples 1 or 2 above. Within 1-7 days the symptoms of the hypothyroidism are alleviated. Thyroid hormone levels return to substantially normal levels. Alleviation of the hypothyroidism persists for at least about 2 months to about 6 months.

Example 4

Treatment of Hypothyroidism With Botulinum Toxin Type B

A 52 year old female is diagnosed with hypothyroidism. Between about 10^{-3} U/kg and about 35 U/kg of a botulinum toxin type B preparation (for example between about 1000 units and about 20,000 units of a botulinum type B

preparation) is injected directly into the thyroid, using one of the techniques set forth in Examples 1 or 2 above. Within 1-7 days the symptoms of the hypothyroidism are alleviated. Thyroid hormone levels return to substantially normal levels. Alleviation of the thyroid disorder persists for at least about 2 months to about 6 months.

Example 5
Treatment of Hypothyroidism With Botulinum Toxin Type C

A 58 year old female is diagnosed with hypothyroidism. Between about 10^{-3} U/kg and about 35 U/kg of a botulinum toxin type C preparation (for example between about 10 units and about 10,000 units of a botulinum type B preparation) is injected directly into the thyroid, using one of the techniques set forth in Examples 1 or 2 above. Within 1-7 days the symptoms of the hypothyroidism are alleviated. Thyroid hormone levels return to substantially normal levels. Alleviation of the thyroid disorder persists for at least about 2 months to about 6 months.

Example 6
Treatment of Hypothyroidism With Botulinum Toxin Type D

A 56 year old obese female is diagnosed with hypothyroidism. Between about 10^{-3} U/kg and about 35 U/kg of a botulinum toxin type D preparation (for example between about 10 units and about 10,000 units of a botulinum type D preparation) is injected directly into the thyroid, using one of the techniques set forth in Examples 1 or 2 above. Within 1-7 days the symptoms of the hypothyroidism are alleviated. Thyroid hormone levels return to substantially normal levels. Alleviation of the thyroid disorder persists for at least about 2 months to about 6 months.

Example 7
Treatment of Hypothyroidism With Botulinum Toxin Type E

A 61 year old female is diagnosed with hypothyroidism. Between about
5 10^{-3} U/kg and about 35 U/kg of a botulinum toxin type E preparation (for example
between about 10 units and about 10,000 units of a botulinum type E
preparation) is injected directly into the thyroid, using one of the techniques set
forth in Examples 1 or 2 above. Within 1-7 days the symptoms of the
hypothyroidism are alleviated. Thyroid hormone levels return to substantially
10 normal levels. Alleviation of the thyroid disorder persists for at least about 2
months to about 6 months.

Example 8
Treatment of Hypothyroidism With Botulinum Toxin Type F

15 A 52 year old male is diagnosed with hypothyroidism. Between about 10^{-3}
U/kg and about 35 U/kg of a botulinum toxin type F preparation (for example
between about 10 units and about 10,000 units of a botulinum type F
preparation) is injected directly into the thyroid, using one of the techniques set
20 forth in Examples 1 or 2 above. Within 1-7 days the symptoms of the
hypothyroidism are alleviated. Thyroid hormone levels return to substantially
normal levels. Alleviation of the thyroid disorder persists for at least about 2
months to about 6 months.

Example 9
Treatment of Hypothyroidism With Botulinum Toxin Type G

25 A 59 year old female is diagnosed with hypothyroidism. Between about
 10^{-3} U/kg and about 35 U/kg of a botulinum toxin type G preparation (for example
30 between about 10 units and about 10,000 units of a botulinum type G
preparation) is injected directly into the thyroid, using one of the techniques set
forth in Examples 1 or 2 above. Within 1-7 days the symptoms of the
hypothyroidism are alleviated. Thyroid hormone levels return to substantially

normal levels. Alleviation of the thyroid disorder persists for at least about 2 months to about 6 months.

Example 10

Treatment of Hyperthyroidism With Botulinum Toxin Type A

A 27 year old female presents with symptoms and signs of hyperthyroidism, including thyrotoxicosis and bilateral ophthalmopathy. Thyroid function tests confirmed the diagnosis of hyperthyroidism. Thyroid scintigraphy shows an enlarged gland and indicates thyroid hyperplasia. A trial with atropine reduced the thyroid hormone level. Between about 10^{-3} U/kg and about 35 U/kg of a botulinum toxin type A preparation (for example between about 10 units and about 300 units of BOTOX®) is injected directly into the cervical ganglia as follows. A percutaneous procedure is carried out using an anterior approach with the patient in a supine position using a very thin needle (22 Gauge) with computed tomography needle guidance to reach the cervical ganglia. Within 1-7 days the symptoms of the hyperthyroidism are alleviated. Thyroid hormone levels return to normal (are lowered). Alleviation of the a thyroid disorder persists for at least about 2 months to about 6 months.

Example 11

Treatment of Hyperthyroidism With Botulinum Toxin Types B-G

A 62 year old female is diagnosed with hyperthyroidism. Between about 10^{-3} U/kg and about 35 U/kg of a botulinum toxin type B, C, D, E, F or G preparation (for example between about 10 units and about 20,000 units of a botulinum toxin type B, C, D, E, F or G preparation) is injected directly into the cervical ganglia as follows. A percutaneous procedure is carried out using an anterior approach with the patient in a supine position using a very thin needle (22 Gauge) with computed tomography needle guidance to reach the cervical ganglia. Within 1-7 days the symptoms of the hyperthyroidism are alleviated. Thyroid hormone

levels return to normal (are lowered). Alleviation of the thyroid disorder persists for at least about 2 to about 6 months.

Example 12

Treatment of Calcium Metabolism Disorders With Botulinum Toxin Types A-G

A 28 year old female is diagnosed with hypercalcemia. Between about 10^{-3} U/kg and about 35 U/kg of a botulinum toxin type A, B, C, D, E, F or G preparation (for example between about 10 units and about 200 units of a botulinum toxin type A preparation) is injected directly into the thyroid using one of the techniques set forth in Examples 1 or 2 above. Within 1-7 days the symptoms of the hypercalcemia are alleviated. Plasma calcium levels return to substantially normal levels. Alleviation of the hypercalcemia persists for at least about 2 months to about 6 months.

Additionally, to treat hypocalcemia between about 10^{-3} U/kg and about 35 U/kg of a botulinum toxin type A, B, C, D, E, F or G preparation (for example between about 10 units and about 200 units of a botulinum toxin type A preparation) is injected directly into the cervical ganglia as follows. A percutaneous procedure is carried out using an anterior approach with the patient in a supine position using a very thin needle (22 Gauge) with computed tomography needle guidance to reach the cervical ganglia. Within 1-7 days the symptoms of the hypocalcemia are alleviated. Plasma calcium levels return to normal (are increased). Alleviation of the hypocalcemia persists for at least about 2 to about 6 months.

Methods according to the invention disclosed herein has many advantages, including the following:

(1) the invention renders unnecessary many surgical procedures for effective treatment of a thyroid disorder.

(2) systemic drug effects can be avoided by direct local application of a neurotoxin according to the present invention.

(3) the ameliorative effects of the present invention can persist, on average, from about 2 months to about 6 months from a single local administration of a neurotoxin as set forth herein.

Although the present invention has been described in detail with regard to certain preferred methods, other embodiments, versions, and modifications within the scope of the present invention are possible. For example, a wide variety of neurotoxins can be effectively used in the methods of the present invention. Additionally, the present invention includes local thyroid administration methods wherein two or more neurotoxins, such as two or more botulinum toxins, are administered concurrently or consecutively. For example, botulinum toxin type A can be administered until a loss of clinical response or neutralizing antibodies develop, followed by administration of botulinum toxin type E.

Alternately, a combination of any two or more of the botulinum serotypes A-G can be locally administered to control the onset and duration of the desired therapeutic result. Furthermore, non-neurotoxin compounds can be administered prior to, concurrently with or subsequent to administration of the neurotoxin to provide adjunct effect such as enhanced or a more rapid onset of denervation before the neurotoxin, such as a botulinum toxin, begins to exert its therapeutic effect.

My invention also includes within its scope the use of a neurotoxin, such as a botulinum toxin, in the preparation of a medicament for the treatment of a thyroid disorder by local administration of the neurotoxin.

Accordingly, the spirit and scope of the following claims should not be limited to the descriptions of the preferred embodiments set forth above.

I claim:

1. A method for treating a thyroid disorder, the method comprising the step of administration of a neurotoxin to a patient, thereby treating a thyroid disorder.

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2. The method of claim 1, wherein the neurotoxin is administered to the thyroid of the patient.

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3. The method of claim 2, wherein the thyroid disorder treated is hypothyroidism.

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4. The method of claim 1, wherein the neurotoxin is administered to a sympathetic ganglion which innervates the thyroid.

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5. The method of claim 4, wherein the thyroid disorder treated is hyperthyroidism.

25

6. The method of claim 1, wherein the neurotoxin is administered in an amount of between about 10^{-3} U/kg and about 35 U/kg.

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7. The method of claim 1, wherein the neurotoxin is made by a Clostridial bacterium.

8. The method of claim 1, wherein the neurotoxin is a botulinum toxin.

9. The method of claim 1, wherein the botulinum toxin is selected from the
5 group consisting of botulinum toxin types A, B, C₁, D, E, F and G.

10. The method of claim 1, wherein the neurotoxin is botulinum toxin type A.

10 11. A method for treating a thyroid disorder, the method comprising the step
of administration of a therapeutically effective amount of a botulinum toxin to a
patient, thereby treating a thyroid disorder.

15 12. The method of claim 11, wherein the botulinum toxin is selected from the
group consisting of botulinum toxin types A, B, C₁, D, E, F and G.

20 13. A method for treating hypothyroidism, the method comprising the step of
local administration to a thyroid of a therapeutically effective amount of a
botulinum toxin, thereby increasing a deficient thyroid hormone secretion from a
thyroid cell and treating hypothyroidism.

25 14. The method of claim 13, wherein the botulinum toxin is selected from the
group consisting of botulinum toxin types A, B, C₁, D, E, F and G.

15. A method for treating hyperthyroidism, the method comprising the step of local administration to a sympathetic ganglion which innervates a thyroid cell of a therapeutically effective amount of a botulinum toxin, thereby reducing an excessive thyroid hormone secretion from the thyroid cell and treating hyperthyroidism.

16. The method of claim 15, wherein the botulinum toxin is selected from the group consisting of botulinum toxin types A, B, C₁, D, E, F and G.

17. A method for treating hypercalcemia, the method comprising the step of local administration to a thyroid C cell of a therapeutically effective amount of a botulinum toxin, thereby increasing a deficient calcitonin secretion from a thyroid C cell and treating hypercalcemia.

18. The method of claim 17, wherein the botulinum toxin is selected from the group consisting of botulinum toxin types A, B, C₁, D, E, F and G.

19. A method for treating hypocalcemia, the method comprising the step of local administration to a sympathetic ganglion which innervates a thyroid C cell of a therapeutically effective amount of a botulinum toxin, thereby decreasing an excessive calcitonin secretion from the thyroid C cell and treating hypocalcemia.

20. The method of claim 19, wherein the botulinum toxin is selected from the group consisting of botulinum toxin types A, B, C₁, D, E, F and G.

ABSTRACT

A method for treating hypothyroidism by local administration of a neurotoxin, such as a botulinum toxin, to a thyroid, thereby reducing an inhibitory effect upon thyroid hormone secretion. A method for treating hyperthyroidism by local
5 administration of a neurotoxin, such as a botulinum toxin, to a sympathetic ganglion which innervates the thyroid, thereby reducing a stimulatory effect upon thyroid hormone secretion. Methods for treating calcium metabolism disorders by local administration of a neurotoxin to modulate calcitonin secretion are also
10 disclosed.

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE
DECLARATION AND POWER OF ATTORNEY

As the inventor named below, I declare that:

This is a declaration and power of attorney to accompany the patent application for the invention entitled "METHOD FOR TREATING THYROID DISORDERS", filed herewith as a 47 page specification, including 20 claims. My residence, post office address, and citizenship are as stated below next to my name. I believe that I am the original, first and sole inventor of the subject matter which is claimed and for which a patent is sought on the invention entitled METHOD FOR TREATING THYROID DISORDERS. The specification of the invention is attached hereto.

I hereby further declare that I have reviewed and understood the contents of the above identified specification, including the claims. I acknowledge the duty to disclose information which is material to the examination of this application in accordance with 37 C.F.R. section 1.56.

As the inventor and applicant for letters patent, I hereby appoint: Stephen Donovan, registration number 33,433; Martin Voet, registration number 25,802; Robert J. Baran, registration number 25,806, and; Carlos Fisher, registration number 36,510, as the attorneys to prosecute this application and to transact all business in the Patent and Trademark Office related thereto.

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I hereby declare that: all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; any and all statements made herein were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment or both, under Section 1001 of Title 18 of the United States Code, and that any such willful false statements may jeopardize the validity of the application or any patent issued thereon.

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